Beneficial Effects of Quercetin on Obesity and Diabetes

Leixuri Aguirre, Noemi Arias, M. Teresa Macarulla, Ana Gracia and Maria P. Portillo*

Group Nutrition and Obesity, Dpt. Nutrition and Food Sciences, Faculty of Pharmacy, University of the Basque Country, Paseo de la Universidad, 7. 01006 Vitoria (Spain), RETIC PREDIMED, Instituto de Salud Carlos III, Spain

Abstract: Scientific research is constantly looking for new molecules that could be used as dietary functional ingredients in the fight against obesity and diabetes, two pathologies highly prevalent in Western societies. In this context, flavonoids represent a group of molecules of increasing interest. The major flavonoid is Quercetin, which belongs to the class called flavonols and is mainly found in apples, tea, onions, nuts, berries, cauliflower, cabbage and many other foods. It exhibits a wide range of biological functions including anticarcenogenic, anti-inflammatory and antiviral; it also inhibits lipid peroxidation, platelet aggregation and capillary permeability. This review focuses on the main effects of Quercetin on obesity and diabetes. The mechanisms of action explaining the effects of Quercetin on these two metabolic disturbances are also considered. Good perspectives have been opened for Quercetin, according to the results obtained either in cell cultures or in animal models. Nevertheless, further studies are needed to better characterize the mechanisms of action underlying the beneficial effects of this flavonoid on these pathologies. Moreover, the body fat-lowering effect and the improvement of glucose homeostasis need to be confirmed in humans. Animal studies have consistently failed to demonstrate adverse effects caused by Quercetin. In contrast, due to inhibitory effect of Quercetin in cytochrome P450, interactions with drugs can be taken into account when they are administered at the same time than Quercetin.

Keywords: Quercetin, obesity, diabetes, insulin resistance, cell cultures and animal models.

INTRODUCTION

Flavonoids belong to a group of natural substances which have a variable phenolic structure and are found in fruits, vegetables, tea and wine. These natural foods were known for their beneficial effects on health long before flavonoids were isolated as the effective compounds. Flavonoids have a basic chemical structure of diphenylpropanes (C6-C3-C6) and are most often found attached to sugar (glycosides), but they can be aglycones, as Quercetin is, consisting of 3 rings and 5 hydroxil groups (Fig. 1). Average daily intake of polyphenols varies in the range of 10-100 mg depending of eating habits [1]. Table 1 shows the foods in which Quercetin is mainly found [2].

It has been reported that Quercetin exhibits a wide range of biological functions including anticarcinogenic, antiinflammatory, antiviral activities. Moreover, it inhibits lipid peroxidation, platelet aggregation and capillary permeability. However, a major concern about this molecule is its poor oral bioavailability, which primarily depends on sugar moiety [3]. Most Quercetin is present in plants as hydrophilic glycosides that are not easily absorbed directly. After hydrolysis of the glycosides, the absorption of Quercetin aglycone is estimated to be as high as 65-81% [4].

This review focuses on the main effects of Quercetin on obesity and diabetes, a metabolic condition associated very often to obesity (Tables 2-5). The mechanisms of

*Address correspondence to this author at the Dpt. Nutrición y Bromatología, Facultad de Farmacia, Paseo de la Universidad, 7, 01006 Vitoria, España; Tel: +34-945-013067; Fax: +34-945-013014; E-mail: mariapuy.portillo@ehu.es

Table 1. Amount of Quercetin in Selected Food (Mangels et al., 1993)

Food	Quercetin (mg/100 g)
Brocoli, raw	2.8
Carrots, raw	0.4
Celery, raw	3.5
Cocoa powder	20.1
Cranberrries, raw	14.0
Kale, raw	5.1
Looseleaf lettuce raw	2.0
Lingonberries, raw	11.3
Onions raw	22.6
Ripe tomatoes	0.5

Fig. (1). Structure of Quercetin.

Table 2. In Vitro Studies Showing the Effects of Quercetin on Adipocytes

Authors/Year	Cell Culture Type	Dose	Effect	Mechanism
Kuppusamy & Das (1992)	Adipocytes from epididymal adipose tissue of male Wistar rats (180-230 g)	30 -50 μΜ	Stimulation of lipolysis.	Inhibition of PDE.
Motoyashiki et al. (1996)	Adipocytes from epididymal adipose tissue of male Wistar rats (200-220 g)	100 - 500 μΜ	Inhibition of the Vanadate- increasing effect on LPL activity	
Hsu &Yen (2006)	3T3-L1 Adipocytes	50 -250 μM	Induction of apoptosis.	 Decrease expression in PARP, Bcl-2, Bax, and Bak proteins Increase expression and activity of caspase 3
Ohkoshi <i>et al.</i> (2007)	Adipocytes from subcutaneous and visceral adipose tissue of female (C5BL/6J) mice (5 weeks old)	100 μΜ		Stimulation of lipolysis of visceral adipose tissue.
Ahn et al. (2008)	3T3-L1 Adipocytes	10-100 μΜ	Induction of adipogenesis.	Decrease in SRBP-1, C/EBPα and PPARγ mRNA expression
			■ Inhibition of <i>de novo</i> lipogenesis	 Decrease in SRBP-1 and FAS mRNA Increase in ACC phosphorylation
Yang <i>et al.</i> (2008)	3T3-L1 mouse embryo fibroblasts	12.5-100 μΜ	■ Increase in apoptosis	

Table 3. In Vivo Studies Showing the Effects of Quercetin on Body Weight and Body Fat

Authors/Year	Animal model	Dose	Time	Effect	Mechanism
Steward <i>et al</i> . (2008)	Male C57BL/6J mice	8 g/kg diet	3 and 8weeks	No effects on body weight or body fat.	Transient increase in energy expenditure at 3 weeks that disappears at 8 weeks
Rivera <i>et al.</i> (2008)	Male Zucker rat (13 weeks old)	10 mg/kg BW/d	10 weeks	Decrease in body weight	
Elberg <i>et al.</i> (2009)	Male and female with a BMI 25-35kg/m ²	150 mg/d	6 weeks	No effect on body weight	
Liang <i>et al</i> . (2009)	Male and female C57BL/6J mice (6 weeks old)	66 mg/kg BW/d	4 weeks	 Protection against body weight gain induced by HF feeding. 	
Kobori <i>et al</i> . (2010)	Male C57/BL6J mice (5 weeks)	5 g/kg diet	20 weeks	Decrease in body weightDecrease in visceral and hepatic fat.	
Wein <i>et al</i> . (2010)	Male Wistar rats (280 g)	25 mg/kg BW/d	4 weeks	Decrease in plasmatic TGNo effect on body weight	Decrease in PPARγ mRNA in adipose tissue
Lai <i>et al</i> . (2011)	Ovariectomized rats (12 weeks old)	20 -625 mg/kg	8 weeks	Decrease in body weight and adiposity	 Increase in lipolysis Increase in oxidation of fatty acids Decrease in adipogenesis.
Kim <i>et al</i> . (2011)	Male Sprague-Dawley (5 weeks old)	100 mg/kg BW/d	7 weeks	No effects on body weight	

Table 4. In Vitro Studies Showing the Effect of Quercetin on Glucose Utilization

Authors/Year	Cell Culture Type	Dose	Effect	Mechanism
Elberg & Shechter (1995)	Rat adipocytes	100 μΜ	Inhibition of the insulin effect on adipose tissue	Inhibition of protein tyrosine kinase
Strobel <i>et al</i> . (2005)	Adipocytes of epididymal tissue of Wistar rat	100 μΜ	Inhibition of methylglucose uptake	 Inhibition of GLUT4 in adipocytes
Kwon et al. (2007)	Caco-2E cells	10- 200 μΜ	Inhibition of glucose uptake	Inhibition of GLUT2
Manzano & Williamson (2010)	Caco-2 cells	31 μΜ	■ Inhibition of glucose uptake	Inhibition GLUT2 and SGLT1
Youl et al. (2010)	INS-1β-cell line and pancreatic cells of male Wistar rats	20 μΜ	Potentiation of both glucose and glibencamide –induced insulin secretion and glucose induced insulin secretion. Protection of β-cells against oxidative damage	 Phosphorylation of ERK1/2 Protection against oxidative damage induced by H₂O₂
Eid <i>et al</i> . (2010)	C2C12 murine skeletal myoblast H4IIE murine hepatocytes	50 μΜ	Enhancement of glucose uptake	Stimulation of AMPK pathway
Torres-Piedra et al. (2010)	HEK 293 cells	10 μΜ		■ Inhibition of 11β-HSD1

Table 5. In Vivo Studies Showing the Effect of Quercetin on Diabetes

Authors/Year	Animal model	Dose	Time	Effect	Mechanism
Vessal et al. (2003)	Male Sprague-Dawley STZ induced diabetes rats (200-220 g)	10 and 15 mg/kg BW/d	10 days	 Decrease in the fasting plasma glucose Improvement of glucose tolerance test Regeneration of the pancreatic islets 	 Increase in hepatic glucokinase activity Increase in the number of pancreatic islets
Coskun et al. (2005)	Male Wistar STZ induced diabetes rats (200-250 g)	15 mg/kg BW/d	4 weeks	 Decrease in serum glucose and increase in insulin in serum Protective effects against oxidative damage Preservation of pancreatic β-cell integrity 	 Decrease in MDA and NO (decreasing lipid peroxidation) Increase in antioxidant enzymes activity: SOD, GSHPx, CAT
Rivera <i>et al</i> . (2008)	Obese Zucker rats (13 weeks old)	2 and 10 mg/kg BW/d	10 weeks	Decrease in hyperinsulinemia	
Kobori <i>et al</i> . (2009)	Male BALB/c STZ induced diabetes mice (7 weeks old)	0.1 and 0.5% diet	2 weeks	 Alleviation of diabetic symptoms and liver injury Decrease in plasma glucose 	Inhibition of Cdkn1a expression in pancreas
Steward <i>et al</i> . (2009)	Male C57BL/6J mice (6 weeks old)	0.8% diet	3 and 8 weeks	 At 3 weeks there is a insulin resistance At 8 weeks the insulin resistance disappears 	Inhibition of insulin- dependent activation of PI-3K
Romero <i>et al.</i> (2010)	SHR and Wistar Kyoto rats (24 weeks old)	10 mg/kg BW/d	4 weeks	No effects on serum glucose and insulin levels and insulin resistance.	
Wein et al. (2010)	Male Wistar rats (280 g)	25mg/kg BW/d	4 weeks	Decrease in hyperinsulinemia	 Increase in adiponectin levels Decrease in PPARγ mRNA in adipose tissue
Kim <i>et al</i> . (2011)	Male C57BL/ksJ-db/db mice (5 weeks old)	0.08% diet	7 weeks	Decrease in fasting plasma glucose levels and glycated hemoglobin	Decrease the intestinal maltose activity
Jung et al. (2011)	Male Sprague-Dawley (8 weeks old)	0-0.1% diet	8 weeks	■ No effect	

action explaining the effects of Quercetin on these two metabolic disturbances are also considered and scheduled in Figs (2 and 3).

QUERCETIN AND OBESITY

Obesity, defined as an excess of adipose tissue when body mass index is $\geq 30 \text{ kg/m}^2$, is due to an imbalance between energy intake and energy expenditure. Obesity is a

major health problem in the industrialized world and it has reached epidemic proportions globally. The World Health Organization estimates that there are more than 1 billion overweight adults, of which at least 300 millions are obese [5]. Moreover, its prevalence is likely to increase as a result of changes in lifestyle, decreased physical activity, and socioeconomic development, among others. In addition, obesity is a complex multi-factorial and chronic disease that

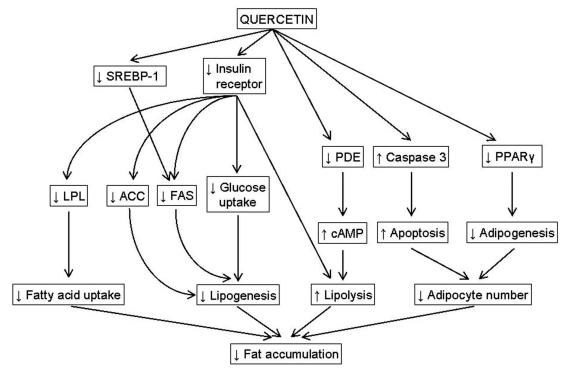


Fig. (2). Proposed mechanisms for body-fat lowering effects of Quercetin in adipose tissue.

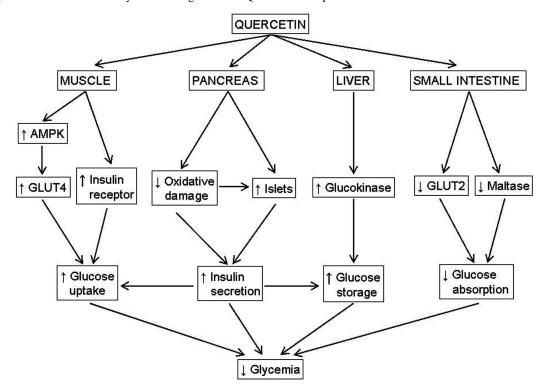


Fig. (3). Proposed mechanisms for anti-diabetic effects of Quercetin.

is considered to be a risk factor for the genesis or development of various diseases including hypertension, type 2 diabetes, coronary heart disease, cancer, respiratory complications and osteoarthritis. Despite current intensive efforts to reduce obesity by diet, exercise, education, drug therapies and surgery, an effective long-term solution to this problem has yet to be provided.

Scientific research is constantly looking for new molecules that could be used as dietary functional ingredients in the fight against overweight and obesity; as in the case of flavonoids.

In Vitro Studies

The first studies addressing the potential anti-obesity action of Quercetin were carried out in vitro, mainly by using cells from adipose tissue and the 3T3-L1 cell line. The incubation of these cells with Quercetin led to a reduction in triacylglycerol content.

Apart from the indirect effects of Quercetin in lipogenesis via insulin, this flavonoid can also inhibit this metabolic pathway by acting directly on the expression of genes controlling this metabolic route. Ahn et al., [6] showed that Quercetin decreased the expression of Sterol Regulatory Element-Binding Proteins (SREBP)-1 and Fatty Acid Synthase (FAS), and by increasing Acetyl-CoA Carboxilasa (ACC) phosphorylation (Fig. 2).

Moreover, Motoyashiki et al., [7] described Quercetin to be a potent inhibitor of the stimulating effect of vanadate on lipoprotein lipase (LPL) activity. Vanadate shows insulinmimetic effects, such as increases in LPL and suppression of hormone-dependent lipolysis, in isolated rat adipocytes. Consequently, the inhibition of vanadate action leads to inhibition of LPL and thus to the incorporation of fatty acids which circulate as triacylgleyerols in lipoproteins to adipocyte triacylglycerols (Fig. 2).

Besides the effects on metabolic pathway involved in triacylglycerol accumulation, Quercetin can also stimulate lipid mobilization. Kuppusamy & Das [8] described that Quercetin induced a dose- and time-dependent increase in lipolysis, which was synergic with epinephrine-induced lipolysis. This flavonoid produces a competitive phosphodiesterase (PDE) inhibition (Fig. 2). The competitive nature of the kinetics suggests that Quercetin could compete with cAMP for the same binding sites in the adipocyte PDE, thus increasing the concentration of cAMP, the molecule that activates protein kinase A (PKA), which in turn activates the hormone sensitive lipase (HSL).

The mechanisms of action described above justify the reduction in adipocyte triacylglycerol content induced by Quercetin. In addition, this flavonoid can also reduce the number of adipocytes, either by decreasing adipogenesis or increasing apoptosis (Fig. 2). Ahn et al., [6] demonstrated that Quercetin attenuated adipogenesis as a consequence of the decrease in the expression of CCAAT/Enhancer Binding Protein α (C/EBPα), and Peroxisome Proliferator-Activated Receptor γ (PPAR γ).

Hsu and Yen [9] described an increase in adipocyte apoptosis due to a down-regulation of PARP (PARP is a protein that has several roles in cellular processes, most notably in DNA repair and programmed cell death), Bcl-2 proteins (apoptosis regulator proteins), Bax, and Bak proteins. The activation of caspase-3 and 9, as well as the inhibition of adenosine monophosphate-activated protein kinase (AMPK) pathway, also play a role in the induction of apoptosis [6].

In Vivo Studies

The first study carried out with animals was reported by Steward et al. [10]. In this study C57BL/6J mice were fed on a high-fat diet (HFD) supplemented with Quercetin (0.8%) for 3 and 8 weeks. Dietary supplementation with this molecule produced a transient (3 weeks) increase in energy expenditure (measured by indirect calorimetry), which was not detected after 8 weeks. A decrease in circulating Quercetin concentration between 3 and 8 weeks suggested a metabolic adaptation. Moreover, Quercetin, at the levels provided, was effective in reducing circulating markers of inflammation (IFN γ , TNF α , IL1 and IL4) after 8 weeks of treatment.

In another study, Rivera et al., [11] analysed the effect of a chronic administration of Quercetin (2 or 20 mg/kg body weight/d) in obese Zucker rats (a model of genetic obesity). In good accordance with Steward et al., [10], final body weight was decreased. Both doses of Quercetin improved dyslipidemia, hypertension, and hyperinsulinemia, but only the high dose produced antiinflammatory effects in visceral adipose tissue.

Using C57BL/6J mice several authors have found reductions in body fat induced by Quercetin treatment. Liang et al. [12] reported that mice fed on a HFD supplemented with Quercetin (66 mg/kg body weight/d) were protected against weight gain induced by the diet. Moreover, Ohkoshi et al. [13] showed a decrease in visceral and subcutaneous adipose tissue. Kobori et al., [14] reported that chronic dietary intake of Quercetin reduced body weight gain, as well as visceral and liver fat accumulation, and improved systemic parameters related to metabolic syndrome (hyperglycemia, hyperinsulinemia and dyslipidemia), probably by decreasing oxidative stress and increasing PPARα expression. Moreover, Quercetin suppressed the expression of PPARy and CD36, as well as SREBP-1c and its target fatty acid synthase (FAS) in the liver. The reduction in PPARy expression suggests a reduction in adipogenesis, because genes involved in this process are controlled by this transcriptional factor. On the other hand, the reduction in SREBP expression, as well as the gene of the lipogenic enzyme FAS, suggest a reduction in the *de novo* lipogenesis in liver. Consequently, a reduced amount of triacylglycerols in plasma, coming from liver and available for adipose tissue uptake, can be suspected.

Nevertheless, not all the published studies have found positive effects of Quercetin on body weight and fat. In an experiment carried out with db/db mice treated with Quercetin (100 mg/kg of body weight/d) for 7 weeks the flavonoid led to a reduction in plasma glucose without changes in body weight [15]. In another experiment performed in Wistar rats treated with 25 mg of Quercetin/kg body weight/d and fed on HFD, body weight also remained unchanged [16].

Very little information has been reported concerning the effects of Quercetin on humans. The only published study shows that nutritional status (body weight, waist circumference, fat mass and fat-free mass) remained unchanged in subjects showing a body mass index between 25 and 35 kg/m², treated with 150 mg/d of Quercetin for 6 weeks [17].

Quercetin Combinations

In recent years, there has been increasing interest from Western Medicine in phytochemical combinations, which have been fundamental in traditional systems of herbal medicine. Combinations of some molecules may synergistically increase their therapeutic activity. Interestingly, in this context Quercetin can help to increase Resveratrol bioavailability. By the mean of in vitro and animal studies it has been shown that the polyphenol Resveratrol shows antiobesity properties [18]. However, a matter of concern is its low bioavailability due to the rapid and extensive metabolism in enterocytes and hepatocytes [19, 20]. It has been proved that some flavonoids are able to inhibit resveratrol metabolism. De Santi and co-workers observed that Quercetin inhibited sulfated metabolites production from Resveratrol, thus increasing the bioavailability of this polyphenol [21].

Apart from the interesting effects of Quercetin on Resveratrol bioavailability, *in vitro* studies have shown that these two polyphenols produce synergic effects. Yang *et al.*, [22] showed that the combination of Resveratrol and Quercetin caused an enhanced increase in apoptosis in 3T3-L1 adipocytes and in maturing preadipocytes inhibition compared to the predicted additive response. These results suggest that Resveratrol and Quercetin, especially in combination, may have a potential to be used for regulating the adipocyte cycle.

In the same line, Lai et al., [23] used a combination of Quercetin with other bioactive molecules (Resveratrol, vitamin D and Genistein) to find a common therapy for both obesity and osteoporosis. Mesenchymal stem cells are the precursors of both adipocytes and osteoblasts. In the aging population, differentiation to adipocytes dominates over the differentiation to osteoblasts in bone marrow. Thus, an inverse relationship exists between adipocytes and osteoblasts in the bone marrow. The above mentioned combination was tested in aged ovariectomized rats, an animal model for postmenopausal bone loss. Rats supplemented 2400 IU/kg vitamin D, 400, 2000 and 1040 mg/kg diet of Resveratrol, Ouercetin and Genistein, respectively, reduced weight gain and adiposity without a change in food intake and increased bone density compared with each molecule administered alone.

OUERCETIN AND DIABETES

Diabetes mellitus is a chronic metabolic disorder which results in disturbances of carbohydrate, protein and lipid metabolism [24] due to either a lack of insulin secretion (type 1) or increased cellular resistance to insulin (type 2). Type 2 diabetes mellitus is one of the world's most common chronic diseases associated with changing lifestyles. It is characterized by hyperglycemia, peripheral resistance to the

insulin action, and eventual destruction of insulin producing β -cells [25].

Under normal physiological conditions, blood glucose levels are tightly regulated by the secretion of insulin by specialized cells in pancreas Langerhans's islets. High blood glucose promotes insulin release from the β -cells of the islets. Insulin stimulates the uptake of glucose from the blood by different tissues such as muscle, kidney and adipose, promotes the storage of glucose in the liver as glycogen, and inhibits lipolysis in adipose tissue. The resulting depletion of blood glucose by the action of insulin in turn promotes the secretion of glucagon from the α -cells in the pancreatic islets, which stimulates glycolysis in the liver and release of glucose back into the blood.

Obesity is a major predisposing factor in the development of Type 2 diabetes. Several hypotheses have been proposed to explain this link and during recent decades a great deal of attention has been given to the large amount of free fatty acids (FFA) received by liver skeletal muscle and pancreas (lipid toxicity) [26]. Moreover, in obese individuals, inflammatory molecules produced by adipose tissue play an important role in producing peripheral insulin resistance, as well as increasing damage to the insulin-producing β -cells [27].

Early treatment and prevention play a pivotal role in reducing burden of diabetes on the population. Lifestyle changes, such as exercising and dietary pattern modifications, are recommended, but these behavioural measures are difficult to maintain in the long term. The benefits of pharmaceutical factors to treat the disease aggressively in its early stages have been recommended, but medications may have unwanted side effects. In this context, flavonoids, among which Quercetin is one of the most commonly found in foods, has been reported to improve diabetic status [28].

In Vitro Studies

Studies carried out in cultured cells have shown that one of the mechanisms of action by which Quercetin improves glycaemic control is the reduction of intestinal glucose absorption at the level of glucose transporters (GLUT) (Fig. 3).

Kwon *et al.*, [29] showed a robust inhibition of Quercetin in glucose and fructose transport by GLUT2 in Caco-2E intestinal cells. However, the two other major intestinal sugar transporters, GLUT5 and SGLT1, were unaffected by this flavonoid. Glucose transport was inhibited by Quercetin in a dose-dependent fashion and was complete at 200 μM Quercetin dose.

In the same research line, Manzano & Williamson [30] studied the effect of strawberry and apples juices, containing 87 and 41 μ M Quercetin respectively, on uptake and apical to basolateral transport of glucose using Caco-2 intestinal cells. Substantial inhibition of both uptake and transport was induced by extracts from both juices. The inhibition of GLUT2 was greater than that of SGLT1.

It has been also postulated that Quercetin blocks tyrosine kinase. Phosphorylation of the specific region of the β -subunit in insulin receptor (including Tyr-1158, Tyr-1161 and Tyr-1162) correlates with receptor tyrosine kinase

activation and the propagation of the biological actions of the hormone [31]. The inhibition of protein tyrosine kinase was also observed by Elberg et al., [32].

Nevertheless, the effect of Quercetin in protein tyrosine kinase is controversial. Thus, Strobel et al., [33] reported that the inhibitory effect of Quercetin on glucose uptake was due to a direct action on the transporter GLUT4, rather than to an effect on cellular protein-tyrosine kinases.

Other authors have described that beneficial effects of Quercetin are due to its effects on the pancreas. Youl et al. [34], using the insulin-secreting cell line INS-1 β , determined the effects of Quercetin (20 µM) on glucose- or glibenclamide-induced insulin secretion, as well as on β-cell dysfunctions induced by hydrogen peroxide (H₂O₂). They observed that Quercetin potentiated both glucose and glibenclamide-induced insulin secretion. This effect was mediated by ERK1/2, a pathway that has been shown to participate in the regulation of glucose induced insulin secretion [35]. In order to confirm this effect in a more physiological model, the authors also used rat isolated islets of Langherans and similar results were obtained. Moreover, a protection against oxidative damage induced by H₂O₂ in β cells was found (Fig. 3).

Another target for Quercetin seems to be AMPK pathway. The study carried out by Eid et al., [36] was aimed to elucidate the mechanism of action of the berries of Vaccinum vitis-idaea, traditionally used for the treatment of diabetes in several cultures throughout the world [37]. The authors analysed the effects of ten compounds present in berries on the glucose uptake in the C2C12 murine skeletal myoblasts and H4IIE murine hepatocytes. They found that Quercetin-3-O-glycoside and Quercetin aglycone were the two most active compounds in glucose uptake. This effect was insulin independent and seems to be mediated by AMPK, which facilitates the translocation of GLUT4 transporter [38] (Fig. 3).

Finally, Torres-Piedra et al., [39] in a study with HEK 293 cells suggested that the 11β-hydroxysteriod dehydrogenase type 1 (11β-HSD1), which mediates glucocorticoid hormone action in human liver, adipose tissues and pancreatic β cells, plays a role in the positive effects of Quercetin on glucose homeostasis. These authors demonstrated that Quercetin induced a 27% inhibition of this enzyme at a concentration of 10 µM.

In Vivo Studies

Several studies have been carried out in diabetic animal models. Kim et al., [15] observed a reduction in serum glucose levels and blood glycated haemoglobin in C57BL/KsJdb-db mice fed for 7 weeks on a diet supplemented with 0.08 % Quercetin, due to the inhibition of small intestine maltase activity, without changes in serum insulin levels (Fig. 3).

Vessal et al., [40] analysed the effects of Quercetin administrated intraperitonally (10 or 15 mg/kg body weight/d) for 10 days in control and streptozoticin-induced (STZ) type 1 diabetes rats. Although Quercetin had no effect on plasma glucose level in control animals, it significantly decreased this parameter in diabetic rats, in a dose dependent manner. Glucose tolerance curves of the diabetic treated animals were very similar to those of control rats. The increase of glucokinase activity induced by Quercetin plays an important role in this effect. This enzyme phosphorylates glucose and converts it into glucose-6-phosphate, a metabolite destined to glycogen synthesis (Fig. 3). Moreover, a significant increase in the number of pancreatic islets was seen in both normoglycemic and diabetic rats treated with Quercetin.

Coskun *et al.*, [41] evaluated the possible protective effect of Quercetin against STZ-induced damage in β cells. This flavonoid partially avoided the increase in serum glucose by decrease in serum insulin concentrations in these type 1 diabetic rats. These effects were due to the fact that Quercetin partially prevented degeneration of β -cells (Fig. 3).

Kobori et al., [42] proved that Quercetin included in the diet (0.1-0.5%) led to the recovery of cell proliferation in mice showing type 1 diabetes induced by STZ. This beneficial effect was mediated by the inhibition of Cdkn1a, which regulates cell division by arresting the cell cycle.

Rivera et al., [43] analysed the effects of chronic administration of two doses of Quercetin on metabolic syndrome, including insulin resistance, in genetically obese Zucker rats. The supplementation was carried during 10 weeks with 2 or 10 mg/kg body weight/d. Both doses of Quercetin improved insulin resistance. No information concerning the mechanism of action underlying this effect was provided in this manuscript.

Wein et al., [16], in a study carried out on male Wistar rats fed on a HFD supplemented with 25 mg Quercetin/kg body weight/d, showed an increase in adiponectin expression in white adipose tissue and its circulating concentration, despite an inhibition of PPARy expression. The authors concluded that the effects of Quercetin on adiponectin were PPARy independent. Taking into account that adiponectin is an adipokine that facilitates insulin action, the authors proposed that the increase in adiponectin levels was involved in the improvement in insulin sensitivity induced by Quercetin.

Although, as described in this review, a great number of papers in the literature have reported positive effects of Quercetin on diabetes, specifically in Type 1 diabetic and genetically diabetic animals, there are also other studies that show either no effects or negative effects in other animal models of diabetes and/or insulin resistance.

Steward et al., [44] induced insulin resistance in rats by feeding animals on a HFD. These authors observed that Quercetin, at a dose of 0.8% in the diet exacerbated dietinduced insulin resistance at 3 weeks. After 8 weeks, insulin resistance in the Quercetin supplemented group was not worse, when compared with animal on HFD alone. These results suggest that the inhibitory effect of insulin signalling at 3 weeks was further eliminated by an adaptive increase hepatic metabolism and/or excretion of the compound between 3-8 weeks. At the doses used, Quercetin inhibited insulin-dependent activation of PI-3K.

To compare the benefits between supplementation with Quercetin and a food rich in Quercetin, Jung et al., [45] carried out an experiment with onion peel extract (OPE) in rats showing Type 2 diabetes. Animals were fed on HFD supplemented with either 1% OPE (containing 0.1% of Quercetin) or 0.1% of Quercetin alone for 8 weeks. While

OPE administration led to significantly improved oral glucose tolerance, Quercetin alone did not have this effect.

Romero et al., [46] carried out a study in normotensive Wistar Kyoto (WKY) rats and in their counterparts, spontaneously hypertensive (SHR) rats In WKY the administration of Quercetin at 10 mg/kg body weight/d for 4 days did not alter serum glucose and insulin levels or insulin resistance. By contrast, in SHR rats, which show increased glucose and insulin levels, as well as insulin resistance when compared with the normotensive counterparts, Quercetin significantly increased fasting serum glucose. A weak non-significant increase in the homeostatic model assessment of insulin resistance was also noticed, indicating that there is a subtle negative effect of Quercetin on glucose metabolism.

Toxicity of Quercetin

In order to set out the potential applications of a biomolecules it is important to know about their safety. In this context, the first toxicity studies related to Quercetin were published in 1970. Bjeldanes & Chang [47] found a positive mutagenic activity in the most standard strains of *Salmonella*. This mutagenic activity observed in bacterial test system was confirmed in eukaryotic cells, including yeast cells, at relatively high concentrations [48]. Additionally, in hamster and mouse cells, as well as in human lymphocytes, Quercetin exposure induced chromosomal aberrations, DNA single strand breaks and micronucleus formation [49].

The results of Quercetin-related mutagenicity/genotoxicity observed *in vitro* have not been confirmed by *in vivo* experiments. After oral administration to mice and rats, Quercetin did not induce any significant change in several mutagenicity/genotoxicity endpoints in somatic cells, in comparison with untreated controls [50-52]. Ambrose *et al.*, [53] carried out an acute toxicity experiment in rabbits. A single intravenous dose of Quercetin (100-150 mg/kg body weight) was injected and no symptoms of toxicity were reported.

Quercetin has also been studied for its potential carcinogenicity in numerous long-term experimental animal studies, the majority of which indicated no evidences of significantly increased incidences of neoplasm formation related to its oral administration [53, 54, 55].

No variations were observed in a number of other standard toxicological parameters, clinical chemistry and organ weights in Swiss mice administered Quercetin at doses of 30, 300, 3000 mg/kg body weight/day for a period of 28 days in comparison with a control group [56]. On the other hand, Quercetin is a strong antioxidant, which may have prooxidant properties under certain conditions. *In vivo*, several protective mechanisms act in concert to limit its potential to act a pro-oxidant/genotoxicant [57].

Taking into account that Quercetin is mainly excreted in urine, a study devoted to analyze potential toxicity in the renal system was carried out by the National Toxicology Program. An increased severity of chronic nephropathy, hyperplasia, and neoplasia of the renal tubular epithelium (causing primarily benign tumors of the renal tubular epithelium) was demonstrated in male, but not in female F344/N rats exposed to 40000 ppm Quercetin in the diet (\approx 2000 mg/kg body weight/day). At lower levels, 1000 ppm

(\approx 50 mg/kg bodyweight/day) and 10000 ppm (\approx 500 mg/kg body weight/day), no statistically significant adverse effects (at p \leq 0.01) were reported [58, 59].

Finally, in the early 1980s, several studies reported the effects of flavonoids on the activity of hepatic cytochrome P450 (P450) enzymes [60]. Since then, the ability of flavonoids to inhibit isoforms of CYP450 has been extensively confirmed [61]. Cytochrome P450 (CYP) monooxygenases are probably the most important enzymes in hepatic drug metabolism, which is crucial for the elimination of many therapeutic drugs [62]. It has been published that Quercetin is a potent inhibitor of CYP2C8 [63, 64], CYP2C9 [65] and CYP3A4 [66] enzymes of the cytochrome P450 at concentrations lower than 50 μM .

The activity of this group of enzymes could determine the patient's response to drug therapy. The interactions between drugs and bioactive molecules could frequently arise when drugs are co-administered. The clinical significance of such interactions also depends on the disposition and toxicity profile of the drug being administered. In the case of cyclosporin, digoxin, indinavir, midazolam, simvastatin and some 1,4-dihydropyridine calcium antagonists, the inhibition of the cytochrome P450 (due to Quercetin intake) could carry out an elevated plasma concentrations with the toxic effects due to its accumulation in the organism [67, 68].

CONCLUSIONS

In the search for new molecules that could be used as dietary functional ingredients in obesity and diabetes, good perspectives have opened up for Quercetin, according to the results obtained either in cell cultures or in animal models. Nevertheless, further studies are needed to better characterize the mechanisms of action underlying the beneficial effects of this flavonoid on these pathologies. Moreover, the body fat-lowering effect and the improvement of glucose homeostasis need to be confirmed in humans.

Animal studies have consistently failed to demonstrate adverse effects caused by Quercetin. In contrast, due to inhibitory effect of Quercetin in cytochrome P450, interactions with drugs can be taken into account when they are administered at the same time than Quercetin.

CONFLICT OF INTEREST

None declared.

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ABBREVIATIONS

ACC = Acetyl-CoA carboxilasa

AMPK = Adenosine monophosphate-activated protein

kinase

FAS = Fatty Acid Synthase

GLUT = Glucose transporter

HFD = High-fat diet

LPL = Lipoprotein lipase

OPE = Onion peel extract

PDE = Phosphodiesterase

PPAR γ = Peroxisome Proliferator-Activated Receptors γ

SHR = Spontaneously hypertensive rats

SREBP = Sterol Regulatory Element-Binding Proteins

STZ = Streptozotocin

WKY = Wistar Kyoto rats

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