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RESEARCH ARTICLE

Synthesis, Characterization and Antifungal Assessment of Optically Active Bis-organotin Compounds Derived from (S)-BINOL Diesters

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Abstract:

Background:

Organotin(IV) derivatives have appeared recently as potential biologically active metallopharmaceuticals exhibiting a variety of therapeutic activities. Hence, it is important to study the synthesis of new organotin compounds with low toxicity that may be of pharmacological interest.

Objectives:

This study focuses on the synthesis of new bis-stannylated derivatives with *C*₂ symmetry that could be tested as antifungal agents against two clinical important fungal species, *Cryptococcus neoformans* and *Candida albicans*.

Methods:

The radical addition of triorganotin hydrides (R₃SnH) and diorganotin chlorohydrides (R₂ClSnH) to bis- α,β -unsaturated diesters derived from (S)-BINOL led to the corresponding new bis-stannylated derivatives with *C*₂ symmetry. Nine pure organotin compounds were synthesized with defined stereochemistry. Four of them were enantiomerically pure and four were diastereoisomeric mixtures.

Results:

All new organotin compounds were fully characterized, those with phenyl ligands bonded to tin were the most active compounds against both the strains (*Cryptococcus neoformans* and *Candida albicans*), with activity parameters of IC₅₀ close to those of the reference drug (amphotericin B).

Conclusion:

Nine pure organotin compounds with *C*₂ symmetry were synthesized with defined stereochemistry and their antifungal properties were tested against two clinical important fungi with IC values close to those of the reference drug. The structure-containing preferably two or three phenyl groups joined to the tin atom were highly active against both the strains compared with those possessing tri-*n*-butyl groups.

Keywords: *C*₂ symmetry, Bis-stannylated (S)-BINOL derivatives, Bis-chlorostannanes, Radical hydrostannation, *Cryptococcus neoformans*, *Cryptococcus neoformans*.

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1. INTRODUCTION

Organotin compounds have been used for many years in different applications, for example as catalysts, heat and light stabilizers, additives of PVC, antifouling, antimold and germicidal agents [1 - 5]. After near 170 years of their disco-

very, Tin(IV) organic compounds are one of the most important concerning organometallic chemistry researches [6 - 10]. Their toxicity depends on the alkyl group, decreasing with the increase in the alkyl chain length [11]. In the last fifteen years, organotin(IV) derivatives have appeared also as potential biologically active metallopharmaceuticals exhibiting anti-tumor, antibacterial, anti-inflammatory and antimicrobial activity [12, 13]. Considering the limitations caused by its toxicity, it is important to develop new organotin compounds

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that could be safer and more effective for chemotherapeutic uses.

Taking into account that bis-chlorostannanes $\text{Ph}_2\text{ClSn-R-SnClPh}_2$ show interesting biological activities [14], we present here the synthesis and antifungal assessment of optically active organotin compounds derived from (S)-BINOL as part of our research work on the behavior of C_2 symmetry diol diesters in radical hydrostannations [15]. The aim of these studies was to determine the scope and limitations of the change in the structural core of the chiral diol over the stereoselectivity in the radical tandem cyclohydrostannation, which lead to eleven-membered macrodiolides [16 - 18]. The starting materials for building these cycles are open chain systems with two carbon-carbon double bonds activated for radical addition by electron withdrawing substituents like the ester group, as shown in Fig. (1). The chiral cores employed in these studies were TADDOL (I) and (1*R*,12*R*)-9,10-dihydro-9,10-ethaneanthracene-11,12-dimethanol (II), both C_2 symmetry diols. Now, in the present study, we added (S)-BINOL (III).

1,1'-Bi-2-naphthol (BINOL) is an axially chiral organic compound that can be found as a racemic mixture or in its two atropisomeric forms: *M* or (*R*) and *P* or (*S*) (Fig. 2). Since the chirality axis is not affected in the esterification reactions, the (S)-BINOL derivatives retain such configuration and optical purity.

The internal relative spatial arrangement of bioactive molecules is crucial regarding their ability to interact favorably with active and allosteric sites. Although since 1990, single enantiomers have clearly dominated over racemates in the Drug Discovery and Regulatory Agency approval trends [19], it is surprising that biologically active atropisomers have not been extensively covered in the literature, despite their prevalence and importance to the pharmaceutical industry. There are several well-documented atropisomeric compounds with demonstrated utility as effective drugs. Among them, it is worth mentioning that the glycopeptide *P*-isomer vancomycin is often used as a drug of penicillin-resistant bacterial infections. It is produced by the soil bacteria *Amycolatopsis*

orientalis, and exhibits broad-spectrum activity against a number of gram-positive bacteria [20]. Another example is gossypol that is produced by *Gossypium hirsutum*, which has been shown to have possible applications as an antifertility and anticancer agent through binding to antiapoptotic protein Bcl-X L [21]. It has recently been shown that the active atropisomer of gossypol is that of the *M*-(-)-enantiomer [22]. As an antecedent of antifungal activity, the atropisomeric flavans myristinins, isolated from *Myristica cinnamomea* fruits, showed antifungal activity against *Candida albicans* with IC_{50} values ranging from 5.9 to 8.8 $\mu\text{g/mL}$ [23].

In the last decades, fungi have emerged as a major cause of human morbidity and mortality, mainly among the immunocompromised and seriously ill-hospitalized patients [24]. Most of the mycoses-related deaths are associated with the spp. *C. albicans* and *Cryptococcus neoformans* and although there are several antifungal drugs in clinical use to fight against this opportunistic fungal spp, the fungal infections remain very difficult to eradicate. *C. albicans* is among the most common cause of opportunistic fungal infections in immunocompromised hosts. *C. neoformans* is the most frequent cause of meningitis and is one of the most important

HIV-related fatal opportunistic mycoses, which has killed more than 650,000 immunocompromised patients worldwide up to date [25]. Besides, *C. neoformans* remains an important life-threatening complication, particularly for immunocompromised patients who have undergone transplantation of solid organs and therefore, new compounds acting against this fungus are highly welcome [26]. Although the incidence of the disease tends to decline in countries with highly active antiretroviral therapy, the outcome of infection is influenced by a variety of factors including the antifungal resistance and new strategies including new structural types with anti-cryptococcal activity are highly welcome [27].

In this work, the synthesized atropisomers (S)-3, (S)-4, (S)-8, (S)-9 and the diastereomeric mixtures 5a-d, 6a-d, 10a-d, 11a-d were tested as antifungal agents against the two mentioned clinical important fungal species, *C. neoformans* and *C. albicans*.

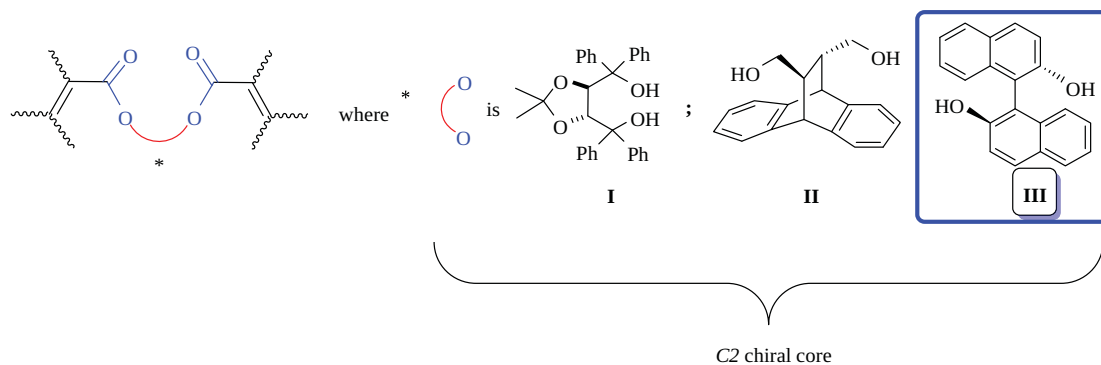


Fig. (1). System of two activated carbon-carbon double bonds linked by a C_2 chiral core.

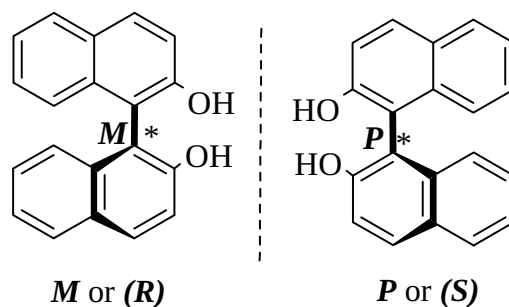


Fig. (2). *M* or (*R*) and *P* or (*S*) BINOL atropoisomers.

2. MATERIALS AND METHODS

Unless otherwise noted, all the reagents were purchased in analytical reagent grade from commercial suppliers and used without purification. All the reactions were carried out under inert atmosphere. The solvents used were distilled and dried in accordance with standard procedures. Di-*n*-butyl- and diphenyltin dihydride were obtained by the reduction of the corresponding dichloride with lithium aluminum hydride [28] and the starting BINOL unsaturated diesters were prepared as described previously [15]. Thin layer chromatography was performed on Merck silica gel 60 F254 plates and visualization was accomplished with UV light and/or 5% ethanol solution of phosphomolybdic acid. Silica gel (Merck, 230–400 mesh) was used for column chromatography. Melting points were recorded on a Büchi Melting Point B-545 instrument and are uncorrected. NMR spectra were recorded on a Bruker Avance 300 Multinuclear instrument, using CDCl_3 as a solvent; chemical shifts (δ) were reported in ppm with respect to TMS in ^1H NMR and ^{13}C NMR, and with respect to Me_4Sn in the case of ^{119}Sn NMR spectra. Data were reported as follows: chemical shift, multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, quin = quintuplet, sex = sextet and br = broad), coupling constant (*J* values in Hz) and integration. Chemical shifts (δ) are reported in ppm and coupling constants (*J*) are in Hz. Infrared spectra were recorded with a Nicolet Nexus 470 FT spectrometer. Optical rotations were measured on a Polar L-P, IBZ Messtechnik polarimeter at 589 nm. Compounds described here were characterized by ^1H , ^{13}C and DEPT-NMR. Elemental analyses (C, H) were performed in an Exeter analytical, Model CE440 Analyzer.

2.1. General Procedure for the Addition of Triorganotin Hydrides to BINOL Unsaturated Diesters Initiated by AIBN at 75°C

Diester (1 mmol), triorganotin hydride (2.4 mmol) and a catalytic amount of AIBN were stirred in dry toluene (24 mL) at 75°C for 1 hour. The reaction was monitored by TLC and IR spectroscopy. The solvent was then distilled off under reduced pressure. The crude product was thus obtained directly purified by column chromatography using silica gel.

2.1.1. (*S*)-1,1'-binaphthalene-2,2'-diyl-bis[3-(tri-*n*-butylstannyl)propanoate] (*S*)-3

The product (*S*)-3 was eluted with hexane/ethyl acetate (98:2) as a yellow oil in a 67% yield. $[\alpha]_{\text{D}}^{25}$ -11.33 (*c* = 0.45,

CHCl_3). ^1H NMR (300 MHz, CDCl_3) δ : 0.71-0.88 (m, 6 x CH_2 , 6 x CH_3 , 30H), 0.86-1.00 (m, 6 x CH_2 , 12H), 1.11-1.31 (m, 6 x CH_2 , 12H), 1.40 (t, $^2J_{(\text{H}, \text{Sn})} = 60.57$, Hz, $^3J_{(\text{H}, \text{H})} = 8.71$, 2 x CH_2 , 4H), 2.46 (t, $^3J_{(\text{H}, \text{Sn})} = 11.8$, Hz, $^3J_{(\text{H}, \text{H})} = 8.71$, 2 x CH_2 , 4H), 7.03-7.12 (m, Ar-H, 4H), 7.15-7.37 (m, Ar-H, 6H), 7.77-7.99 (m, Ar-H, 2H). ^{13}C NMR (75.4 MHz, CDCl_3) δ : 2.72 (468.8), 8.83 (323.2), 13.80, 27.39 (56.3), 29.14 (19.5), 31.17 (16.4), 110.82, 117.76, 124.04, 124.20, 127.48, 128.41, 129.45, 131.43, 133.39, 151.74, 173.62 (64.2). ^{119}Sn NMR (CDCl_3) δ : -7.39 ppm. Anal. Calcd. for $\text{C}_{50}\text{H}_{74}\text{O}_4\text{Sn}_2$: C, 61.50, H, 7.64. Found: C, 61.64, H, 7.70.

2.1.2. (*S*)-1,1'-binaphthalene-2,2'-diyl-bis[3-(triphenylestannyl)propanoate] (*S*)-4

The product (*S*)-4 was eluted with hexane/ethyl acetate (90:10) as colourless oil in a 86% yield. $[\alpha]_{\text{D}}^{25}$ -11.09 (*c* = 0.82, CHCl_3). ^1H NMR (300 MHz, CDCl_3) δ : 1.41 (t, $^2J_{(\text{H}, \text{Sn})} = 61.8$, Hz, $^3J_{(\text{H}, \text{H})} = 7.42$ Hz, 2 x CH_2 , 4H), 2.51 (t, $^3J_{(\text{H}, \text{H})} = 7.42$ Hz, 2 x CH_2 , 4H), 7.23 (d, $^3J_{(\text{H}, \text{H})} = 8.9$ Hz, Ar-H, 2H), 7.36-7.45 (m, Ar-H, 4H), 7.46-7.58 (m, Ar-H, 20H), 7.62-7.65 (m, Ar-H, 12H), 7.97-8.00 (m, Ar-H, 4H). ^{13}C NMR (75.4 MHz, CDCl_3) δ : 5.02 (390.1), 29.34 (18.4), 122.03, 123.44, 125.35, 125.72, 126.13, 126.74, 128.54, 128.85, 128.97, 129.06, 131.48, 133.36, 137.06 (35.3), 137.48, 138.28 (504.5), 146.81, 171.42 (64.9). ^{119}Sn NMR (CDCl_3) δ : -99.05 ppm. Anal. Calcd. for $\text{C}_{62}\text{H}_{50}\text{O}_4\text{Sn}_2$: C, 67.91, H, 4.60. Found: C, 68.05, H, 4.67.

2.1.3. 1,1'-binaphthalene-2,2'-diyl-bis[2-methyl-3-(tri-*n*-butylestannyl)propanoate] (*5a-d*, *5a-b*)

The mixture of *5a-d* was eluted with hexane/ethyl ether (97:3) as a white solid in a 67% yield. By re-chromatography, the diastereomers *5a-b* were eluted with hexane/ethyl ether (96:4) as a white solid in a 34% yield in a 61:39 ratio [d.e.: 22%]. ^1H NMR (300 MHz, CDCl_3) δ : 0.46 (dd, $^3J_{(\text{H}, \text{H})} = 6.9$ Hz, $^2J_{(\text{H}, \text{H})} = 2.3$ Hz, 4 x CH_2 , 8H), 0.60-0.67 (m, 12 x CH_2 , 24H), 0.81 (t, $^3J_{(\text{H}, \text{H})} = 7.3$ Hz, 12 x CH_3 , 36H), 1.11-1.22 (m, 4 CH_3 y 12 CH_2 , 36H), 1.23-1.33 (m, 12 x CH_2 , 24H), 2.23-2.42 (m, 4 x CH, 4H), 7.21-4.23 (m, Ar-H, 8H), 7.28-7.42 (m, Ar-H, 8H), 7.77-7.93 (m, Ar-H, 8H). ^{13}C NMR (75.4 MHz, CDCl_3) δ : 9.39 (309.9), 9.41 (323.6), 12.27 (281.5), 13.84, 19.73 (22.8), 27.32 (55.5), 29.20 (17.0), 37.30 (23.6), 37.33 (20.2), 122.11, 123.81, 123.83, 125.70, 126.27, 126.76, 127.93, 129.30, 129.34, 131.59, 131.61, 133.56, 133.60, 147.02, 175.91, 175.92. ^{119}Sn NMR (CDCl_3) δ : -11.34, -11.82. Anal. Calcd. for

C₅₂H₇₈O₄Sn₂: C, 62.17, H, 7.83. Found: C, 62.32, H, 7.89.

2.1.4. 1,1'-binaphthalene-2,2'-diyl-bis[2-methyl-3-(triphenylstannyl)propanoate] (6a-d)

The mixture of **6a-d** was eluted with hexane/ethyl acetate (95:5) as a white solid in a 84% yield. ¹HNMR (300 MHz, CDCl₃) δ: 0.42-0.60 (m, 8 x CH₃, 24H), 0.84-1.08 (m, 8 x CH₂, 16H), 2.39-2.66 (m, 8 x CH, 8H), 6.92-7.12 (m, Ar-H, 16H), 7.38-7.55 (m, Ar-H, 64H), 7.59-7.70 (m, Ar-H, 56H), 7.84-7.93 (m, Ar-H, 32H). ¹³CNMR (75.4 MHz, CDCl₃) δ: 13.59, 13.62, 13.70, 13.72, 18.29, 35.34, 35.29, 35.27, 35.23, 120.22, 120.25, 120.29, 120.31, 121.72, 121.75, 123.90, 123.94, 124.25, 124.26, 124.36, 124.90, 124.92, 126.13, 126.18, 126.41, 126.39, 126.74, 126.94, 126.99, 127.06, 127.11, 127.13, 129.68, 129.71, 129.73, 131.54, 131.56, 131.61, 131.62, 153.33, 135.71, 136.98, 137.01, 137.03, 137.36, 144.95, 144.09, 145.01, 173.29, 173.34, 173.38, 173.41. ¹¹⁹SnNMR (CDCl₃) δ: -103.24, -103.61, -103.74, -103.91. Anal. Calcd. for C₆₄H₅₄O₄Sn₂: C, 68.36, H, 4.84. Found: C, 68.53, H, 4.94.

2.2. General Procedure for the Addition of di-*n*-butyltin Chlorohydride to BINOL Unsaturated Diesters Initiated with AIBN at 40°C (METHOD A) [17]

Di-*n*-butyltin dichloride (0.18 g, 0.61 mmoles) was dissolved in dry toluene (3.4 mL), and di-*n*-butyltin dihydride (0.12 mL, 0.14 g, 0.61 mmoles) was added. The mixture was stirred for about 30 min, with monitoring the reaction by IR spectroscopy to verify the formation of di-*n*-butyltin chlorohydride. Then, a solution of diester (0.51 mmoles) in dry toluene (5.7 mL) was added with a syringe slowly to the mixture together with a catalytic amount of azo-bis-isobutyronitrile (AIBN) as a radical initiator. The reaction mixture was heated to 40°C and was monitored by TLC and IR spectroscopy. After one hour, the solvent was then distilled off under reduced pressure. The crude product thus obtained was directly purified by column chromatography using silica gel 60.

2.3. General Procedure for the Addition of Diphenyltin Chlorohydride to BINOL Unsaturated Diesters Initiated with AIBN at 40°C (METHOD A) [17]

Diphenyltin dichloride (0.21 g, 0.61 mmoles) was dissolved in dry toluene (1 mL), and diphenyltin dihydride (0.12 mL, 0.17 g, 0.61 mmoles) was added. The mixture was stirred for about 1 hour, monitoring the reaction by IR spectroscopy to verify the formation of diphenyltin chlorohydride. Then, a solution of diester (0.51 mmoles) in dry toluene (5.7 mL) was added with a syringe slowly to the mixture together with a catalytic amount of azo-bis-isobutyronitrile (AIBN) as a radical initiator. The reaction mixture was heated to 40°C and was monitored by TLC and IR spectroscopy. After one hour, the solvent was then distilled off under reduced pressure. The crude product thus obtained was directly purified by column chromatography using silica gel 60.

2.4. Procedure for the Addition of di-*n*-butyltin Chlorohydride to BINOL Dimethacrylate Diesters at -78°C Initiated with Et₃B (METHOD B) [17, 34]

Di-*n*-butyltin dihydride (0.12 mL, 0.14 g, 0.61 mmoles) was added to a solution of di-*n*-butyltin dichloride (0.18 g, 0.61 mmoles) in dry toluene (3.4 mL) at room temperature. After being stirred for 30 min, the formation of the di-*n*-butyltin chlorohydride was verified by IR and the mixture was cooled to -78°C. A solution of diester (**S**)-**1b** (0.21 g, 0.51 mmoles) in dry toluene (5.7 mL) was added with a syringe slowly to the mixture. Immediately, Et₃B (0.10 mmoles, 0.0099 g, 0.011 mL) was added and the mixture was stirred at -78°C for 8 h with monitoring by TLC and IR spectroscopy. The solvent was distilled off under reduced pressure.

2.5. Procedure for the Addition of Diphenyltin Chlorohydride to BINOL Dimethacrylate Diesters at -78°C Initiated with Et₃B (METHOD B) [17, 34]

Diphenyltin dihydride (0.11 mL, 0.16 g, 0.57 mmoles) was added to a solution of diphenyltin dichloride (0.19 g, 0.57 mmoles) in dry toluene (0.5 mL) at room temperature. After being stirred for 1 hour, the formation of diphenyltin chlorohydride was verified by IR and the mixture was cooled down to -78°C. A solution of diester (0.47 mmoles) in dry toluene (4.5 mL) was added with a syringe slowly to the mixture. Immediately, Et₃B (0.094 mmoles, 0.0092 g, 0.010 mL) was added and the mixture was stirred at -78°C for 10 h with monitoring by TLC and IR spectroscopy. The solvent was distilled off under reduced pressure.

2.5.1. (S)-1,1'-binaphthalene-2,2'-diyl-bis[3-(chlorodi-*n*-butylestannyl)propanoate] (S)-8 Method A

The product (**S**)-**8** was eluted with hexane/ethyl acetate (98:2) as a yellow oil in a 65% yield. [α]_D²⁵ +170.80 (c=0.020, CHCl₃). ¹HNMR (300 MHz, CDCl₃) δ: 0.77 (dd, ³J_{(H,H)}} = 7.3 Hz, 2CH₂, 4H), 0.83 (dd, ³J_{(H,H)}} = 7.3 Hz, 2CH₂, 4H), 1.06-1.39 (m, 4 x CH₃, 4 x CH₂, 20H), 1.43-4.56 (m, 4 x CH₂, 8H), 2.22-2.33 (m, 2 x CH₂, 4H), 2.49-2.60 (m, 2 x CH₂, 4H), 7.02-7.33 (m, Ar-H, 6H), 7.43 (dt, ³J_{(H,H)}} = 8.2 Hz, ⁴J_{(H,H)}} = 1.8 Hz, Ar-H, 2H), 7.89 (d, ³J_{(H,H)}} = 8.2 Hz, Ar-H, 2H), 7.43 (d, ³J_{(H,H)}} = 8.2 Hz, Ar-H, 2H). ¹³CNMR (75.4 MHz, CDCl₃) δ: 11.59 (388.6), 13.81, 18.11 (647.0), 18.80, 26.88 (85.1), 27.86 (41.1), 29.96 (27.2), 120.83, 123.06, 126.24, 126.46, 127.40, 128.39, 130.48, 131.92, 133.06, 146.34, 178.77 (24.4). ¹¹⁹SnNMR (CDCl₃) δ: 85.88 ppm. Anal. Calcd. for C₄₂H₅₆Cl₂O₄Sn₂: C, 54.06, H, 6.05. Found: C, 54.24, H, 6.12.

2.5.2. (S)-1,1'-binaphthalene-2,2'-diyl-bis[3-(chlorodiphenylestannyl)propanoate] (S)-9 Method A

The product (**S**)-**9** was eluted with hexane/ethyl acetate (98:2) as a yellow oil in a 95% yield. [α]_D²⁵ +36.32 (c=0.038, CHCl₃). ¹HNMR (300 MHz, CDCl₃) δ: 1.26-1.30 (m, CH₂, 2H), 1.30-1.40 (m, CH₂, 2H), 2.16-2.28 (m, 2 x CH₂, 4H), 6.83-7.07 (m, Ar-H, 8H), 7.21-7.42 (m, Ar-H, 20H), 7.82-7.89 (m, Ar-H, 4H). ¹³CNMR (75.4 MHz, CDCl₃) δ: 13.04 (513.0), 29.38 (32.8), 120.72, 122.57, 125.30, 125.64, 126.38, 127.31, 128.22, 128.58, 129.10, 129.17, 129.65, 130.14, 130.49, 131.66,

132.723, 136.11 (45.3), 146.13, 179.36 (43.6). $^{119}\text{SnNMR}$ (CDCl_3) δ : -52.55. Anal. Calcd. for $\text{C}_{50}\text{H}_{40}\text{Cl}_2\text{O}_4\text{Sn}_2$: C, 59.27, H, 3.98. Found: C, 59.45, H, 4.07.

2.5.3. 1,1'-binaphthalene-2,2'-diyl-bis[3-(chlorodi-n-butyls-tannyl)-2-methyl propanoate] (10a-d) Method A

The mixture of diastereoisomers **10a-d** was isolated by column chromatography using silica gel 60 as the stationary phase eluting as a yellow oil with a mixture of hexane: ethyl acetate (90:10) in a 78% yield. *Method B*. The mixture of diastereoisomers **10a-d** was isolated by column chromatography using silica gel 60 as the stationary phase eluting as a yellow oil with a mixture of hexane: ethyl acetate (90:10) in a 40% yield. $^1\text{HNMR}$ (300 MHz, CDCl_3) δ : 0.18 (d, $^3J_{(\text{H,H})} = 7.2$ Hz, 2 x CH_3 , 6H), 0.30 (d, $^3J_{(\text{H,H})} = 7.2$ Hz, 2 x CH_3 , 6H), 0.59 (d, $^3J_{(\text{H,H})} = 7.2$ Hz, 2 x CH_3 , 6H), 0.76-0.91 (m, 2 CH_3 , 4 x 2 CH_2 , 22H), 1.04-1.84 (m, 4 x 12 CH_2 , 4 x 4 CH_3 , 144H), 2.37-2.91 (m, 4 x 2CH, 8H), 7.04-7.32 (m, Ar-H, 24H), 7.40-7.45 (m, Ar-H, 8H); 7.86-7.97 (m, Ar-H, 16H). $^{13}\text{CNMR}$ (75.4 MHz, CDCl_3) δ : 12.68, 18.33, 18.63, 18.90, 19.00, 20.55, 20.56, 20.73, 20.85, 25.43, 25.57, 26.08, 26.28, 31.27, 31.87, 35.55, 35.62, 35.66, 35.74, 119.65, 121.97, 124.74, 125.32, 126.22, 127.20, 129.07, 130.71, 131.82, 145.12, 180.03, 180.28, 180.35, 180.61. $^{119}\text{SnNMR}$ (CDCl_3) δ : 72.95, 70.09, 69.68, 68.72. Anal. Calcd. for $\text{C}_{44}\text{H}_{60}\text{Cl}_2\text{O}_4\text{Sn}_2$: C, 54.98, H, 6.29. Found: C, 55.12, H, 6.35.

2.5.4. 1,1'-binaphthalene-2,2'-diyl-bis[3-(Chlorodiphenylest-annyl)-2-methyl Propanoate] (11a-d) Method A

The mixture of diastereoisomers **11a-d** was purified by column chromatography using silica gel 60 as the stationary phase eluting as a yellow oil with a mixture of hexane: ethyl acetate (98:2) in a 96% yield. *Method B*. The mixture of diastereoisomers **11a-d** was isolated by column chromatography using silica gel 60 as the stationary phase eluting as a yellow oil with a mixture of hexane: ethyl acetate (98:2) in a 65% yield. $^1\text{HNMR}$ (300 MHz, CDCl_3) δ : 0.04 (d, $^3J_{(\text{H,H})} = 7.2$ Hz, 2 x CH_3 , 6H), 0.19 (d, $^3J_{(\text{H,H})} = 7.2$ Hz, 2 x CH_3 , 6H), 0.28 (d, $^3J_{(\text{H,H})} = 7.2$ Hz, 2 x CH_3 , 6H), 0.44 (d, $^3J_{(\text{H,H})} = 7.2$ Hz, 2 x CH_3 , 6H), 0.89-1.34 (m, 4 x 2 CH_2 , 16H), 2.32-2.78 (m, 4 x 2CH, 8H), 6.61-7.22 (m, Ar-H, 112H), 7.78-7.88 (m, Ar-H, 16H). $^{13}\text{CNMR}$ (75.4 MHz, CDCl_3) δ : 18.78, 19.08, 19.18, 19.32, 23.32, 23.40, 36.45, 36.53, 36.60, 36.73, 118.77, 118.93, 119.00, 120.74, 120.94, 121.28, 123.60, 123.71, 123.78, 123.83, 124.63, 124.73, 124.78, 125.60, 125.70, 126.81, 127.42, 127.85, 128.26, 128.75, 129.91, 129.99, 130.11, 130.99, 131.01, 131.08, 134.41, 144.20, 144.29, 144.31, 144.36, 181.25, 181.63, 181.75, 181.85. $^{119}\text{SnNMR}$ (CDCl_3) δ : -63.69, -66.73, -68.68, -70.39. Anal. Calcd. for $\text{C}_{52}\text{H}_{44}\text{Cl}_2\text{O}_4\text{Sn}_2$: C, 59.98, H, 4.26. Found: C, 60.12, H, 4.34.

2.6. Biology, Antifungal Evaluation, Microorganisms and Media

For the antifungal evaluation, the standardized strains *C. albicans* ATCC 10231 and *C. neoformans* ATCC 32264 from the American Type Culture Collection (ATCC), Rockville, MD, USA were used. Strains were grown on Sabouraud-chloramphenicol agar slants for 48 h at 30°C, maintained on

slopes of Sabouraud-dextrose Agar (SDA, Oxoid), and subcultured every 15 days to prevent pleomorphic transformations. Inocula of the cell or spore suspensions were obtained according to reported procedures and adjusted to 1-5x 10³ cells/spores with colony forming units (CFU)/mL.

2.7. Antifungal Susceptibility Testing

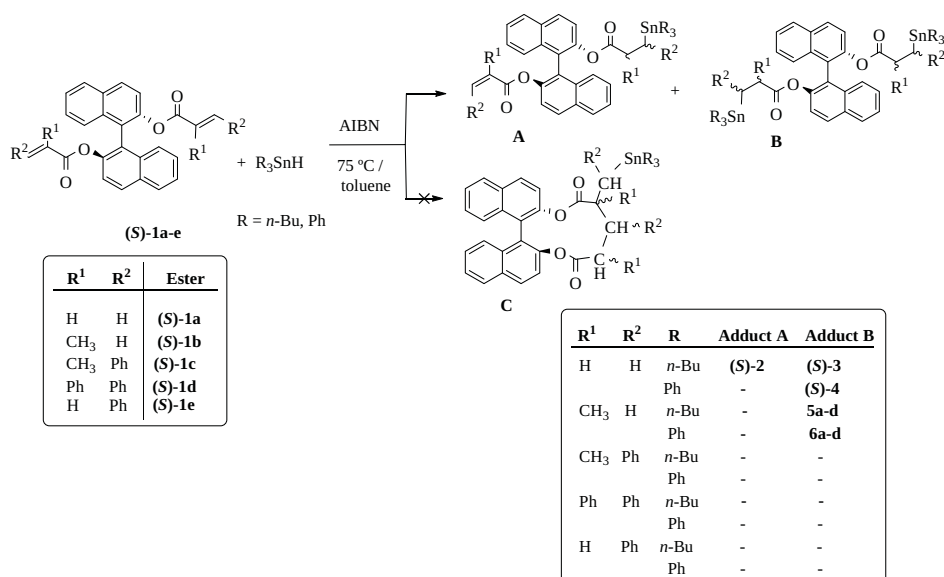
Minimum Inhibitory Concentration (MIC) of each extract or compound was determined by using broth microdilution techniques according to the guidelines of the National Committee for Clinical Laboratory Standards for yeasts (M27-A3). MIC values were determined in RPMI-1640 (Sigma-Aldrich, St Louis, MO, USA), buffered to pH 7.0 with MOPS. Microtiter trays were incubated at 35°C in a moist, dark chamber, and MICs were visually recorded at 48 h. For the assay, stock solutions of pure compounds were two-fold diluted with RPMI from 256 to 3.9 $\mu\text{g/mL}$ (final volume = 100 μL) and a final DMSO concentration $\leq 1\%$. A volume of 100 μL of inoculum suspension was added to each well with the exception of the sterility control where sterile water was added to the well instead. Amphotericin B (Sigma Aldrich) was used as a positive control. Endpoints were defined as the lowest concentration of drug resulting in total inhibition (MIC) of visual growth compared to the growth in the control wells containing no antifungal. The IC_{50} was defined as the lowest concentration of a compound that produced 50% reduction in the growth control (culture media with the microorganism but without the addition of any compound), and was determined spectrophotometrically with the aid of a VERSA Max microplate reader (Molecular Devices, Sunnyvale, CA, USA) and the % of inhibition for each compound at the different concentrations was calculated as follows: 100x (OD (optical density) 405 MTW - OD 405 SCW)/(OD 405 GCW - OD 405 SCW). Tests were conducted in triplicate.

2.8. X-ray Diffraction Analysis

The X-ray data were collected at 293(2) K with a Nonius Kappa-CCD with Mo source diffractometer. The used software for data collection included: APEX3: Bruker (2017). APEX3. Bruker AXS Inc., Madison, Wisconsin, USA; for structure solution SHELXT [29] and for structure refinement SHELXL [30]. All H atoms bonded to carbon were placed with idealized geometry and refined using riding model with C-H = 0.95 Å, $U_{\text{iso}}(\text{H}) = 1.2 U_{\text{eq}}(\text{C})$ for CH. The absolute configuration was determined by the Flack parameter and was assumed to be unchanged during the reaction and was not determined by diffraction methods. The structural data have been deposited with Cambridge Crystallographic Data Centre, CCDC number CCDC 1589534. Crystallographic data of (**S**)-**1e** for the structural analyses have been deposited with the Cambridge Crystallographic Data Centre, CCDC 1589534. Copies of this information may be obtained free of charge from the Director, CCDC, 12 Union Road, Cambridge, CB21EZ, UK, fax: +44 1223 336 033; email deposit@ccdc.cam.ac.uk or www:http://www.ccdc.cam.ac.uk.

3. RESULTS AND DISCUSSION

Based on our previous results [16 - 18], we assumed that the radical hydrostannation of (S)-BINOL unsaturated diesters



Scheme (1). Radical addition of triorganotin hydrides to (*S*)-BINOL unsaturated diesters.

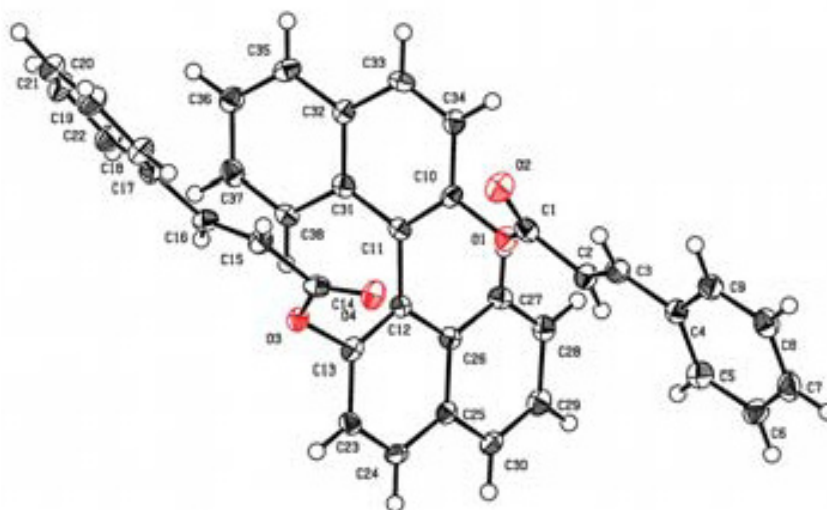


Fig. (3). X-ray structure of diester (*S*)-1e.

(*S*)-1a-e could lead to additional products to one or both olefinic systems (compounds type **A** or **B**, Scheme 1) or the corresponding cycloadducts (compounds type **C**, Scheme 1).

After performing the reaction between (*S*)-1,1'-binaphthyl-2,2'-diyl-diacylates ((*S*)-1a-e) and the corresponding tri-*n*-butyl- and triphenyltin hydrides under already known radical conditions [16 - 18, 31], (anhydrous toluene, 1:1,2 substrate/organotin hydride relation, inert atmosphere, 75°C and AIBN as radical initiator) only mixtures of type **A** and/or **B** adducts were formed when substrates (*S*)-1a and (*S*)-1b were employed. No reaction was observed with (*S*)-1c to (*S*)-1e, and the expected cyclized structure **C** was not observed in any case. The (*S*)-BINOL diester spatial arrangement could explain these results. This molecule, with *C*₂ symmetry, adopts preferably the transoid conformation, where the bulky substituents located in positions 2 and 2' mutually move away. Thus, the olefinic

groups are arranged opposite to the other and, therefore, the formation of the macrocycle is disfavored. This structural arrangement was confirmed through X-ray diffraction analysis of dicinnamate (*S*)-1e which produced suitable crystals that would confirm this approach (Fig. 3). Dicinnamate (*S*)-1e was obtained following the procedure of reference 13. (See Supporting Information for X-Ray diffraction analysis). Crystal structural data was deposited at the Cambridge Crystallographic Data Centre with Deposition number: CCDC 1589534.

Increasing the proportion of organotin hydride, the optimal substrate / R₃SnH relation determined in order to obtain a higher proportion of type **B** bis-trialkyltin addition products was 1:2,4 under the previously mentioned conditions. TLC chromatographic monitoring of the reaction showed that they were completed after one hour. Electronic and steric factors

usually determine the rate of addition to olefins and in both cases, the alkene has the ester group as an activating electron-withdrawing group and the β -carbon is unsubstituted so there is no steric hindrance for the regioselective binding of the trialkylstannyl group to the terminal carbon. However, this last factor would affect diesters (**S-1c** to **S-1e**), which remained unreacted after 10 h. Table 1 summarizes the results obtained in the radical hydrostannylation of the reactive substrates (**S-1a** and **S-1b**).

The reaction with Bu_3SnH with the diester (**S-1a**), gave a 89% mixture of mono- and di-addition products (**S-2** and **S-3**) in a 31:69 proportion respectively which could be separated by chromatography although with low final yield of pure bis-stannylated adduct (**S-3**) (Table 1, entry 1, 67%, Supporting Information, Fig. S1). Increasing the substrate/ Bu_3SnH relation to 1:3 provides a 20:80 mixture of single (**A**) and double addition products (**B**) but in lower yield (63%) from which (**S-3**) was obtained in a 45% yield. Type **A** adduct (**S-2**) could not be isolated in a pure form so it could not be characterized. The same organotin hydride reacted with substrate (**S-1b**) to yield in this case only bis-addition adducts **5a-d**, obtained as a mixture of four diastereomers in relation 50:13:25:12 (Table 1, entry 3, 67%) from which it was possible to separate two of them by re-chromatography, named **5a-b** (34%), $^{119}\text{SnNMR}$ (δ , ppm) -11.34 and -11.82 (see Supporting Information, Fig. S2 and experimental section).

The best yields were observed in the reaction with Ph_3SnH and only type **B** products were obtained. This can be attributed to the higher reactivity and higher hydrogen donating capability of Ph_3SnH relative to Bu_3SnH . (**S-4**) adduct was obtained as a single enantiomer in 86% yield after purification (Table 1, entry 2; Supporting Information, Fig. S3), and **6a-d** were obtained in 84% yield, as a mixture of four diastereomers in 8:19:42:31 relation, that could not be separated (Table 1, entry 4; Supporting Information, Fig. S4). All the obtained double addition products were purified by chromatography and characterized by ^1H , ^{13}C and $^{119}\text{SnNMR}$. In all cases, the diastereomeric composition (% D) was determined from the $^{119}\text{SnNMR}$ spectra of the crude reaction products. Analysis of

^1H and ^{13}C NMR spectral data (Supporting Information, Tables S1-S4 and experimental section) confirms the structure of type **B** adducts (**S-3**), (**S-4**), **5a-b** and **6a-d**, showing in the ^{13}C NMR spectrum only one signal with $^3J_{(\text{Sn,C})}$ coupling constant, corresponding to C=O bond, for each stereoisomer.

Changing the organic ligand by halogen in the organotin hydride determines novel derivatives with the general structure of bis-halodiorganotin adduct and the differentiation of certain molecular properties that would help further studies thereof [32 - 34].

As it is already known, a halogen atom as a ligand bound to tin renders the latter more electropositive, which favors its intramolecular coordination with the carbonyl in β -respective to the ester group through a five-member cycle (Fig. 4) which could determine a particular spectroscopic behavior [35].

Haloorganotin derivatives of this type could be obtained through radical addition of mixed organotin halohydrides R_2SnHX . These highly reactive tin hydrides undergo radical chain reactions at low temperature (-78°C to 40°C) not higher than 40°C in order to prevent their decomposition. Their higher Lewis acidity (in comparison with the most commonly used hydrostannylation reagent Bu_3SnH or Ph_3SnH) leads to much better regio- and stereoselectivities.¹⁷ Considering the formation of an intermediate with intramolecular coordination during the mechanistic steps of the radical halohydrostannylation, this could result in a higher stereoselectivity or differences in the course of the reaction.

We added dialkyltin chlorohydrides R_2SnHCl ($\text{R} = n\text{-Bu}$, Ph) to the (*S*)-BINOL unsaturated diesters (**S-1a** and **S-1b**), and then analyzed whether substantial changes in diastereoselectivity and yield were produced in the products obtained (Scheme 2). The mixed hydrides were prepared by exchange reactions between an equimolecular mixture of R_2SnH_2 and R_2SnCl_2 and used *in-situ* [36]. The halohydrostannations were carried out under inert atmosphere and radical conditions, using AIBN as radical initiator, using 2.4 equivalents of R_2SnHCl in toluene and keeping the mixture at room temperature (Method A) [37].

Table 1. Radical hydrostannylation of (*S*)-1a and (*S*)-1b.

Entry	Diester	R_3SnH	Diester/ R_3SnH Relation	A/ B ^a	$^{119}\text{SnNMR}$ (δ , ppm) ^c				Yield (%) ^d	% D ^e
					(A) ^b	Cpd. N ^o	(B) ^c	Cpd. N ^o		
1	(S)-1a	<i>n</i> - Bu_3SnH	1: 2.4	31/69	-7.08	(S)-2	-7.39	(S)-3	67	100
		<i>n</i> - Bu_3SnH	1: 3	20/80	-7.08	(S)-2	-7.39	(S)-3	45	100
2		Ph_3SnH	1: 2.4	0/100	- ^f	-	-99.05	(S)-4	86	100
3	(S)-1b	<i>n</i> - Bu_3SnH	1: 2.4	0/100	- ^f	-	-11.34	5a-d	67	50
							-11.40			13
							-11.82			25
							-11.86			12
4		Ph_3SnH	1: 2.4	0/100	- ^f	-	-103.38	6a-d	84	8
							-103.73			19
							-104.02			42
							-104.20			31

^a Proportion of mono (A) and di-addition (B) products determined by integration in the $^{119}\text{SnNMR}$ spectrum; ^b Single addition adduct; ^c Double addition adduct; ^d Chemical shift determined in CDCl_3 respect to Me_4Sn ; ^e Yield of the isolated di-addition products; ^f Diastereomeric relationship determined by integration in the $^{119}\text{SnNMR}$ spectrum; ^g Not observed

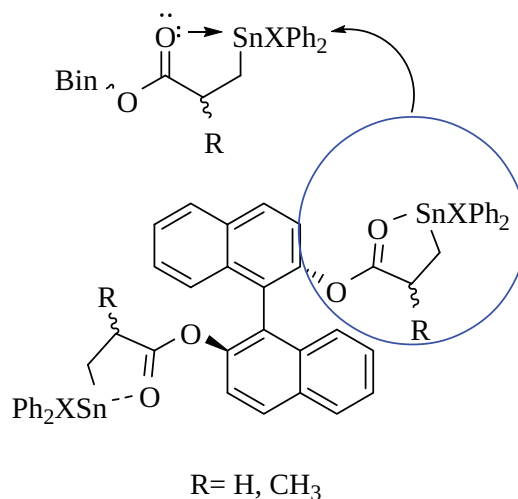
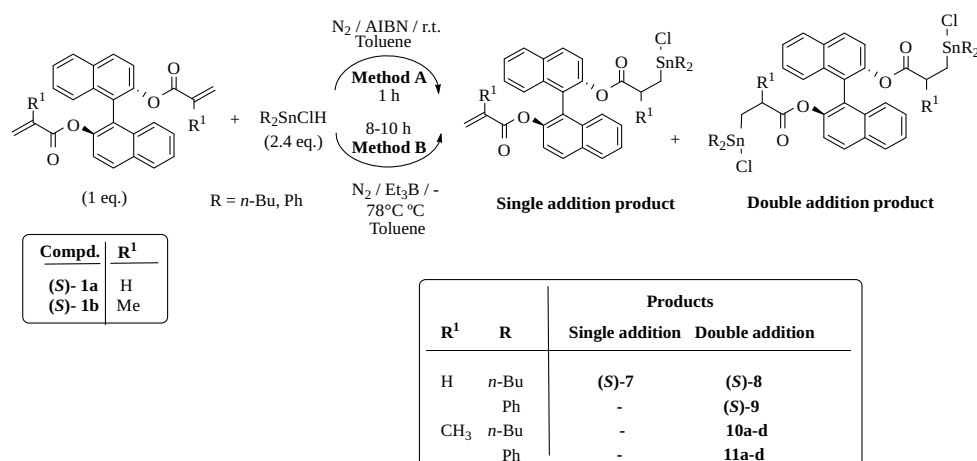


Fig. (4). Five-member cycle intramolecular coordination in β -haloorganotin esters.



Scheme (2). Addition of dialkyltin chlorohydrides R₂SnHCl to the unsaturated diesters (S)-1a and (S)-1b.

Table 2. Radical chlorohydrostannation of 1a and 1b by thermal induction (Method A).

Entry	Diester	-R	S/D Products Relation ^a	¹¹⁹ SnNMR (δ , ppm) ^b			Yield (%) ^c	%D ^f
				(S) ^c	(D) ^d	Cpd. N ^o		
1	(S)-1a	<i>n</i> -Bu	8/92	87.46 (S)-7	85.88	(S)-8	65	100
2		Ph	0/100	- ^g	-52.55	(S)-9	95	100
3	(S)-1b	<i>n</i> -Bu	0/100	- ^g	72.95	10a-d	78	19
70.09					32			
69.68					25			
68.72					24			
4	(S)-1b	Ph	0/100	- ^g	-63.69	11a-d	96	22
-66.73					29			
-68.68					25			
-70.39					24			

^a By integration in the ¹¹⁹SnNMR spectrum; ^b Chemical shift determined in CDCl₃ respect to Me₄Sn;

^c Single addition adduct; ^d Double addition adduct; ^e Yield of the isolated bis-addition product; ^f Diastereomeric relationship determined by integration in the ¹¹⁹SnNMR spectrum of the crude reaction product; ^g No signals are observed.

Except for the addition of *n*-Bu₂SnHCl to (**S**)-**1a** where a single addition product was detected (Table 2, entry 1), the reaction gave exclusively double addition products after 1 h. Yields were almost quantitative for Ph₂SnHCl addition and smaller for *n*-Bu₂SnHCl probably due to its lower reactivity (H donor capability). The diastereoisomeric mixtures **10a-d** and **11a-d** could not be separated and no improvement in diastereoselectivity was observed under these conditions.

Taking into account that the stereoselectivity in radical reactions may be improved by temperature lowering [17, 38], we studied the chlorohydrostannations of **1b** at -78°C using Et₃B as a radical initiator (Method B, Table 3) but without success. We observed longer reaction times, lower yields and no improvement in the diastereomeric relation. Spectroscopic characteristics of this bis-chloroorganotin adducts are reported in the Supporting Information, Tables S5-S8.

All the products obtained, either those enantiomerically pure (**S**)-**3**, (**S**)-**4**, (**S**)-**8**, (**S**)-**9** or the diastereoisomeric mixtures **5a-d**, **6a-d**, **10a-d**, **11a-d** were tested for antifungal activity assays. Despite having separated two diastereoisomers (**5a-b**), the mixture of four (**5a-d**) was used in order to have a uniform comparison criterion to all diastereomeric mixtures.

4. ANTIFUNGAL ACTIVITY ASSAYS

Compounds **S-3**, **S-4**, **S-8**, **S-9**, **5a-d**, **6a-d**, **10a-d** and **11a-d** were tested for antifungal properties against *C. albicans* ATCC 10231 and *C. neoformans* ATCC 32276 (Table 4). To assess antifungal activities, the broth microdilution method M27-A3 for yeasts of the Clinical and Laboratory Standards Institute was used [39].

The percentage of inhibition of each fungus was determined for each compound at two-fold dilutions in the range 250-3.9-μg/mL where MIC and IC₅₀ (the minimum concentration that inhibits 100 and 50% of fungal growth) are shown in the rightmost columns. Amphotericin B (Sigma Aldrich, St Louis, MO, USA) was used as a positive control.

The results obtained in the antifungal assays against *C. albicans* and *C. neoformans* (Table 4) allowed us to observe some trends in the structural-activity relationships: (a) There is

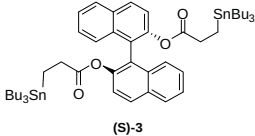
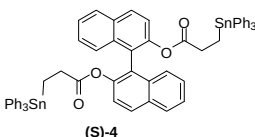
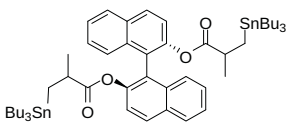
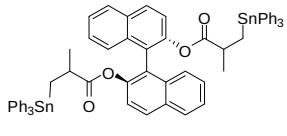
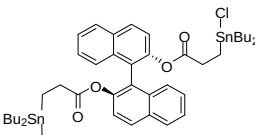
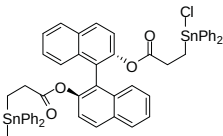
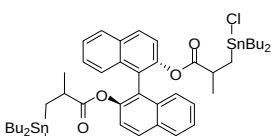
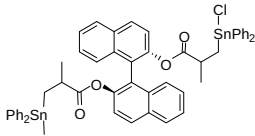
not a clear difference in activity of the compounds against *C. albicans* and *C. neoformans* in the case of (**S**)-**3**, (**S**)-**4**, **6a-d** and **5a-d**. For example, it can be observed that the diastereomers **6a-d** possess an IC₅₀ of 1 μg/mL against both the species, considering that one dilution is the error of the method [40]. Stereoisomers (**S**)-**3**, (**S**)-**4**, and **5a-d** gave similar values of IC₅₀ = 250 μg/mL. However, the behavior of (**S**)-**9**, **11a-d**, (**S**)-**8** and **10a-d** is different. It can be observed that the activity of the enantiomeric form (**S**)-**9**, while is important against both strains, is lower for *C. albicans* than for *C. neoformans* (IC₅₀ = 3.9 μg/mL and 1.9 μg/mL respectively) while in the case of diastereomers **11a-d** and enantiomer (**S**)-**8**, the IC₅₀ values are not so good (31.25 and 15.6 μg/mL; 62.5 and 125 μg/mL). In turn, diastereoisomers **10a-d** showed a very good IC₅₀ value of 2 μg/mL for *C. albicans* but low activity for *C. neoformans* (IC₅₀ = 15.6 μg/mL). (For the sake of clarity see Fig. (5). A and B). (b) If the comparisons are made taking into account different organic ligands on the Sn atom, there are clear differences in the activity: (b1) most compounds with Ph₂Sn-substituents like **6a-d**, with IC₅₀ = 1 μg/mL for both fungi (except for (**S**)-**4** that possesses IC₅₀ = 250 μg/mL) are highly active compared with those possessing *n*-Bu₃Sn- (**5a-d**, (**S**)-**3**, IC₅₀ = 250 and >250 μg/mL respectively for both fungi). (b2) A methyl group in the alpha position (and therefore the introduction of a new chiral center leading to diastereomers) shows that the methyl group does not play a clear role in the antifungal activity. For example, **6a-d** possess higher activity than (**S**)-**4**, whereas **11a-d** show lower activities than (**S**)-**9**. (c) In contrast with point (b), there are clear differences in activity when comparing compounds with different substituents on the Sn atom: (c1) from the compounds containing SnPh₃ (**6a-d** and (**S**)-**4**), **6a-d** is highly active (IC₅₀ values = 1 μg/mL for both fungi) compared with those possessing SnBu₃ ((**S**)-**3** and **5a-d** which showed IC₅₀ = 250 - > 250 μg/mL for both fungi); (c2) When one phenyl of SnPh₃ is changed into Cl leading to SnPh₂Cl ((**S**)-**9** and **11a-d**) changes in the activity are observed: the IC₅₀ of (**S**)-**9** is much better than that observed for (**S**)-**4**. However, when **11a-d** is compared with **6a-d**, the antifungal effect is reversed, from an IC₅₀ = 31.25 for *C. albicans* and 15.6

Table 3. Radical chlorohydrostannation of **1b** at -78°C (Method B).

Entry	-R	¹¹⁹ SnNMR ^a			Yield (%) ^d	Time (hs)	%D ^e
		(S) ^b	(D) ^c	Cpd. N ^o			
1	<i>n</i> -Bu	-	72.95	10a-d	40	8	34
			70.09				16
			69.68				26
			68.72				24
2	Ph	-	-63.69	11a-d	65	10	22
			-66.73				29
			-68.68				17
			-70.39				31

^a Chemical shift determined in CDCl₃ respect to Me₄Sn; ^b Single addition adduct; ^c Double addition adduct; ^d Yield of isolated products; ^e Diastereomeric relationship determined by integration in the ¹¹⁹SnNMR spectrum of the crude reaction product; ^f No signals are observed.

Table 4. Percentages of inhibition of organotin compounds against *C. albicans* ATCC 10231 (*Ca*) and *C. neoformans* ATCC 32264 (*Cn*).

Structure	Fungi	Concentrations ($\mu\text{g/mL}$)										MIC ^a	IC ₅₀ ^a
		250	125	62.5	31.2	15.6	7.8	3.9	2	1	0.5		
Percentages of Inhibition (%)													
 (S)-3	<i>Ca</i>	3.5 \pm 0.8	1.0 \pm 0.7	0.7 \pm 0.1	0.6 \pm 0.0	0.4 \pm 0.1	0	0	0	0	0	>250	>250
	<i>Cn</i>	0	0	0	0	0	0	0	0	0	0	>250	>250
 (S)-4	<i>Ca</i>	53.1 \pm 0.0	28.9 \pm 1.3	15.8 \pm 0.1	5.9 \pm 0.6	4.5 \pm 0.0	2.7 \pm 0.2	2.3 \pm 0.4	0	0	0	>250	250
	<i>Cn</i>	65.9 \pm 0.8	46.6 \pm 0.5	15.8 \pm 1.9	15.0 \pm 0.2	9.7 \pm 0.1	0	0	0	0	0	>250	250
 5a-d	<i>Ca</i>	70.2 \pm 0.1	18.1 \pm 1.5	1.73 \pm 1.8	0.6 \pm 0.9	0	0	0	0	0	>250	250	
	<i>Cn</i>	39.5 \pm 0.8	12.6 \pm 0.2	11.5 \pm 0.2	9.8 \pm 0.2	6.6 \pm 0.2	6.4 \pm 0.8	5.9 \pm 0.2	0	0	0	>250	>250
 6a-d	<i>Ca</i>	100	100	100	100	100	100	100	97.1 \pm 0.0	68.9 \pm 0.2	21.9 \pm 0.8	3.9	1
	<i>Cn</i>	100	100	100	100	100	100	100	100	68.4 \pm 0.5	31.1 \pm 0.5	2	1
 (S)-8	<i>Ca</i>	100	100	96.0 \pm 1.1	33.0 \pm 0.5	29.4 \pm 0.1	5.1 \pm 1.0	3.4 \pm 0.4	0	0	0	125	62.5
	<i>Cn</i>	93.1 \pm 0.8	66.3 \pm 2.0	30.1 \pm 0.3	18.3 \pm 0.9	9.4 \pm 1.3	8.5 \pm 1.0	8.1 \pm 1.7	0	0	0	>250	125
 (S)-9	<i>Ca</i>	100	100	100	100	100	100	100	32.4 \pm 0.6	9.6 \pm 0.1	0	3.9	3.9
	<i>Cn</i>	100	100	100	100	100	100	100	52.0 \pm 1.6	35.7 \pm 0.3	12.6 \pm 1.0	3.9	1.9
 10a-d	<i>Ca</i>	100	100	100	100	100	100	85.7 \pm 0.1	51.4 \pm 1.0	27.5 \pm 0.6	3.8 \pm 0.7	7.8	2
	<i>Cn</i>	100	100	100	100	65.4 \pm 0.6	30.0 \pm 0.3	11.5 \pm 1.1	0	0	0	31.25	15.6
 11a-d	<i>Ca</i>	100	100	93.2 \pm 0.2	51.8 \pm 1.7	10.5 \pm 1.1	3.4 \pm 0.1	0.7 \pm 0.7	0	0	0	125	31.25
	<i>Cn</i>	100	100	97.4 \pm 0.9	87.7 \pm 0.2	49.3 \pm 0.4	22.4 \pm 2.2	7.8 \pm 2.0	0	0	0	125	15.6
Amphotericin B	<i>Ca</i>	100	100	100	100	100	100	100	100	100	100	1.5	0.5
	<i>Cn</i>	100	100	100	100	100	100	100	100	100	100	1.5	0.25

^a in $\mu\text{g/mL}$.

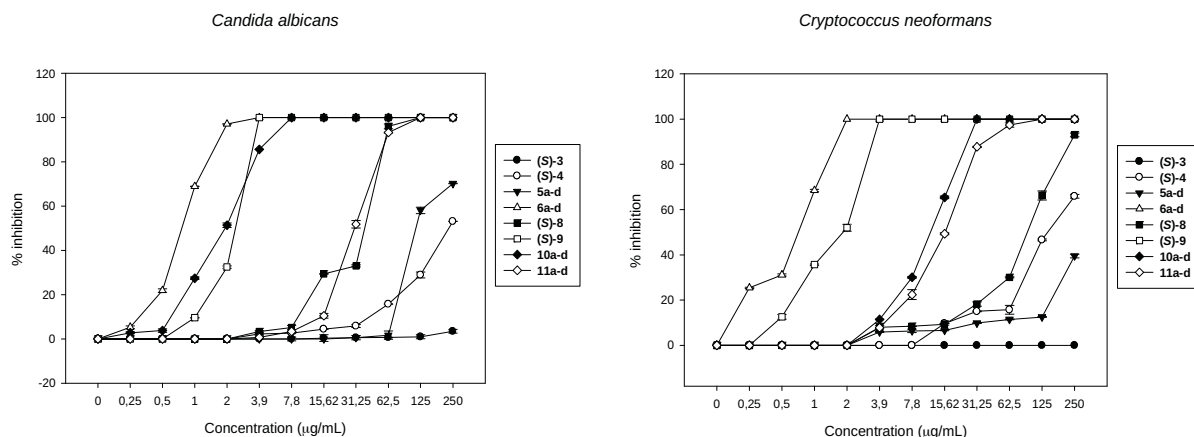


Fig. (5). Comparative curves of the growth inhibition of (A) *C. albicans* ATCC 10231 and (B) *C. neoformans* produced by all tested compounds at different concentrations. Inhibition percentages are the means \pm SD obtained from experiments in triplicate.

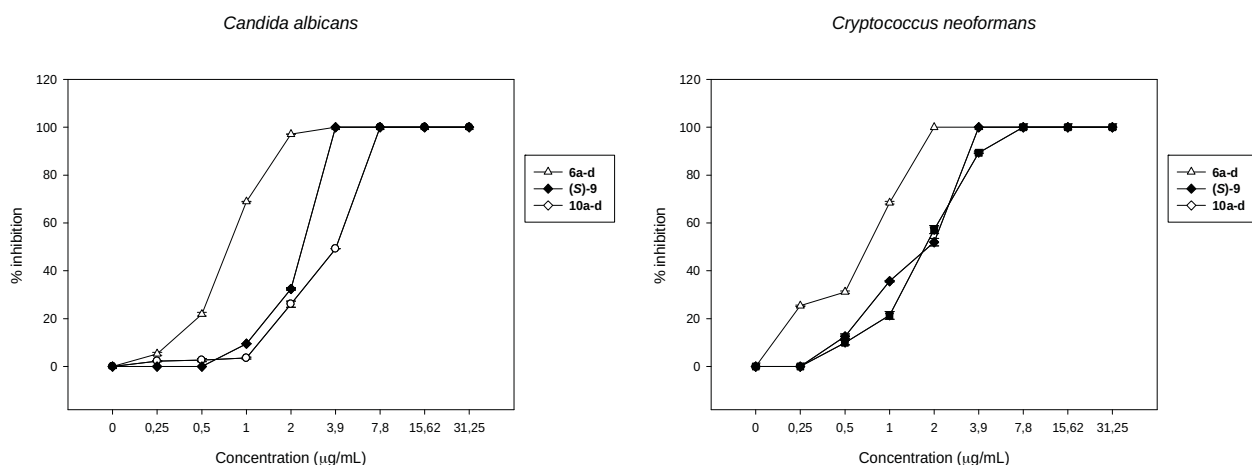


Fig. (6). Comparative curves of the growth inhibition of (A) *C. albicans* ATCC 10231 and (B) *C. neoformans* produced by the most active compounds at concentrations lower than $31.25 \mu\text{g/mL}$. Inhibition percentages are the means \pm SD obtained from experiments in triplicate.

$\mu\text{g/mL}$ for *C. neoformans* (**11a-d**) to an $\text{IC}_{50} = 1 \mu\text{g/mL}$ for **6a-d** in both fungi. (c3) When SnBu_3 in (**S**)-**3** and **5a-d** is changed into SnBu_2Cl , leading to (**S**)-**8** and **10a-d** respectively, the IC_{50} decreases from $\text{IC}_{50} \geq 250$ in (**S**)-**3** to 62.5 - $125 \mu\text{g/mL}$ in (**S**)-**8** and from $250 \mu\text{g/mL}$ in **5a-d** to 2 - 15.6 in **10a-d**, the last one rendering a highly active compound. A comparative curve of the behavior of the most active structures against *C. albicans* and *C. neoformans* can be observed in Figs. (5 and 6).

CONCLUSION

After all these studies it could be assumed that the spatial disposition of the unsaturated ester groups linked to the *C2* chiral core in (*S*)-BINOL diesters (**S**)-**1a-e** determines that the radical addition of organotin hydrides or organotin halo-hydrides leads to double addition adducts. From the nine pure organotin compounds synthesized here with defined stereochemistry, four of them ((**S**)-**3**, (**S**)-**4**, (**S**)-**8** and (**S**)-**9**) are enantiomerically pure and four (**5a-d**, **6a-d**, **10a-d** and **11a-d**)

are diastereoisomeric mixtures. The antifungal properties were tested against two clinical important fungi with the diastereomeric mixture **6a-d**, being the most active against both strains, followed by the enantiomer (**S**)-**9** and the diastereomeric mixture **10a-d**. Interesting enough, the active compounds behave similarly against *C. albicans* and *C. neoformans* and some of them possess IC values close to those of the reference drug. Regarding the most active structural features, these compounds contain preferably two or three phenyl groups joined to the tin atom bonded to an atropisomeric core. Adducts **6a-d**, with SnPh_3 moiety, are highly active against both fungi compared with those possessing SnBu_3 group.

SUPPLEMENTARY MATERIAL

Supplementary material is available on the publisher's website along with the published article.

AVAILABILITY OF DATA AND MATERIALS

The structural data have been deposited with Cambridge Crystallographic Data Centre, CCDC number CCDC 1589534. Crystallographic data of (**S**)-**1e** for the structural analyses have been deposited with the Cambridge Crystallographic Data Centre, CCDC 1589534.

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CONSENT FOR PUBLICATION

Not applicable.

CONFLICT OF INTEREST

The authors declare no conflict of interest, financial or otherwise.

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Declared none.

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